## Alkylation of $\beta$ -(Hydroxymethyl)pyrroles: A New Synthesis of Porphobilinogen and **Other Trisubstituted Pyrroles for Photodynamic Therapy**

## Christine Y. De Leon and Bruce Ganem\*

Department of Chemistry, Baker Laboratory, Cornell University, Ithaca, New York 14853-1301

## Received October 23, 1996

The trisubstituted pyrrole porphobilinogen (PBG; 1 in Scheme 1) is a key intermediate in the biosynthesis of porphyrins and corrins and the central building block for the biosynthesis of hemoglobin, chlorophyll, and many other brightly-colored pigments of life.<sup>1</sup> First isolated<sup>2</sup> and assigned its structure 40 years ago,3,4 PBG has remained a popular synthetic target. Successful total syntheses have been reported from substituted pyrroles,<sup>5</sup> from pyrrole itself,<sup>6</sup> and from azaindole.<sup>7</sup> Yet despite its deceptively simple-looking achiral structure, PBG remains difficult to synthesize in preparatively useful amounts and expensive to buy (>\$10 000 per gram).8

Interest in a practical and efficient route to PBG that could furnish analogs and congeners has been fueled by the emergence of photodynamic therapy (PDT) using synthetic, porphyrin-based pigments to treat systemic cancers, superficial malignancies, and small, localized tumors.<sup>9</sup> PDT relies on the administration of chromophores that sensitize cells to irradiation with visible or near-visible light to generate singlet oxygen. Appropriate sensitizers must show selectivity for the malignant tissue and must be cleared promptly in order to achieve timely treatment regimens. In a major advance, oral administration of 5-aminolevulinic acid, the biosynthetic precursor of PBG, was shown to sensitize tumor cells to light for only a few hours.<sup>10</sup> Since 2-(aminomethyl)pyrroles like 1 can self-assemble nonenzymatically into uroporphinoids quite readily under neutral<sup>11</sup> or acidic<sup>3</sup> conditions, we reasoned that PBG and suitable

(1) Battersby, A. R. Science 1994, 264, 1551.

(2) Westall, R. G. Nature 1952, 170, 614.

(3) Cookson, G. H.; Rimington, C. Biochem. J. 1954, 57, 476.

(4) (a) Raper, R.; Prasad, K. S. N. *Nature* **1955**, *175*, 629. (b) Rimington, C.; Krol, S. *Nature* **1955**, *175*, 630.

(5) (a) Jackson, A. H.; McDonald, D. M.; McDonald, S. F. J. Am. Chem. Soc. 1956, 78, 505. (b) Jackson, A. H.; MacDonald, S. F. Can. J. Chem. 1957, 35, 715. (c) Arsenault, G. P.; MacDonald, S. F. Can J. Chem. 1961, 39, 2043. (d) Plieninger, H.; Hess, P.; Ruppert, J. Chem. Ber. 1968, 101, 240. (e) Battersby, A. R.; Moron, J.; McDonald, E.; Feeney, J. J. Chem. Soc., Chem. Commun. 1972, 920. (f) Kenner, G. W.; Smith, K. M.; Unsworth, J. F. J. Chem. Soc., Chem. Commun. 1973, 43. (g) Battersby, A. R.; Hunt, E.; McDonald, E.; Feeney, J. J. Chem. Soc., Perkin Trans. 1 1973, 2973. (h) Jones, M.; Froussions, C.; Evans, D. A. J. Chem. Soc., Chem. Commun. 1976, 472. (i) Kenner, G. W.; Rimmer, J.; Smith, K. M.; Unsworth, J. F. J. Chem. Soc., Perkin Trans. I 1977, 332. (j) Ufer, G.; Tjoa, S. S.; MacDonald, S. F. Can. J. Chem. 1978, 56, 2437. (k) Adamczyk, M.; Reddy, R. E. Tetrahedron Lett. 1995, 36, 9121; (l) 1996, 37, 2325.

(6) (a) Demopoulos, B. J.; Anderson, H. J.; Loader, C. E.; Faber, K. Can. J. Chem. **1983**, 61, 2415. (b) Faber, K.; Anderson, H. J.; Loader, C. E.; Daley, C. S. Can. J. Chem. **1984**, 62, 1046.

(7) (a) Frydman, B.; Reil, S.; Despuy, M. E.; Rapoport, H. J. Am. Chem. Soc. **1969**, *91*, 2338. (b) Frydman, B.; Buldain, G.; Repetto, J. C. J. Org. Chem. **1973**, *38*, 1824. (c) Valasinas, A.; Levy, E. S.; Frydman, B. J. Org. Chem. **1974**, *39*, 2874. (d) Battersby, A. R.; McDonald, E.; Wurziger, H. K. W.; James, K. J. J. Chem. Soc., Chem. *Commun.* **1975**, 493. (8) Aldrich Chemical Company, 1996/7 Catalog.

(9) Bonnett, R. *Chem. Soc. Rev.* **1995**, *24*, 19. (10) Grant, W. E.; Hopper, C.; MacRobert, A. J.; Speight, P. M.; Bown, S. G. Lancet 1993, 342, 147.

(11) Mauzerall. D. J. Am. Chem. Soc. 1960, 82, 2605.



<sup>a</sup> Key: (a) TsCH(Li)N=C; (b) POCl<sub>3</sub>, DMF; (c) NH<sub>2</sub>OH-EtOH; (d) H<sub>2</sub>, Pd(OH)<sub>2</sub>, HCl, EtOH; (e) NaOEt-EtOH; (f) DIBAL-THF, rt; (g) dimethyl malonate (5 equiv), NaH (5 equiv), 24 h; (h) NaCN (1.5 equiv), DMF, cat. H<sub>2</sub>O, 130-140 °C, 24 h.

analogs might be transformed into porphyrin or corrinlike pigments, either in vivo or in vitro, for PDT.

To implement the first stage of that strategy, we report a short, high-yielding synthesis of PBG itself and of several related 2-(aminomethyl)pyrroles. Key features of our approach include (1) an adaptation of the van Leusen pyrrole construction to the acidic electrophile, diethyl glutaconate, (2) a highly regioselective aminomethylation of the product pyrroloacetic ester, and (3) a new method for the direct alkylation of 3-(hydroxymethyl)pyrroles that accommodates a variety of simple carbon nucleophiles such as malonates, substituted malonates, acetoacetates, and cyanide. Of special interest is the capability of installing acetate, propionate, or larger side chains at the 3-position of the pyrrole nucleus leading to PBG analogs and homologs 2-5.

Simple enones and enoates react with the sodium salt of tosylmethyl isocyanide (TosMIC) to afford 3- or 3,4disubstituted pyrroles.<sup>12</sup> Under those conditions, however, diethyl glutaconate (6) (Scheme 2) underwent deprotonation and self-condensed to form a dimeric tetraester as the exclusive product. To minimize proton transfer and retard dimerization, 6 (1 equiv) was added to the lithium salt of TosMIC [1 equiv, THF, -70 °C, generated using LiN(TMS)<sub>2</sub>] with slow warming to rt. In that fashion, multigram quantities of pyrrole diester 7 could be obtained in 70-80% yield. Formylation of 7 under Vilsmeier-Haack conditions gave the expected aldehyde 8 as the predominant product (8:1 ratio, 96% overall) that, without separation, was converted to the

© 1996 American Chemical Society

<sup>(12)</sup> van Leusen, A. M.; Siderius, H.; Hoogenboom, B. E.; van Leusen, D. Tetrahedron Lett. 1972, 5337.

oxime **9** and hydrogenated over Pd(OH)<sub>2</sub>. Upon basic workup, the desired amino diester **10** cyclized to lactam **11**, which was easily obtained pure by chromatography in 83% overall yield from **8**.

Selective reduction of 11 with DIBAL under controlled conditions furnished (hydroxymethyl)pyrrololactam 12 (86%), a key intermediate in our synthesis. We envisioned that substitution of the OH group in 12 with appropriate carbon nucleophiles might proceed without activation as an acetate or tosylate by a process of elimination–addition involving a  $\beta$ -alkylidenepyrrolenine cation.<sup>13</sup> In fact, when **12** was heated with the sodium salt of dimethyl malonate (5 equiv, DMF, 130 °C, 4 h), diester 13 was obtained in 65% yield. Nucleophilic deesterification and decarboxylation (1.3 equiv NaCN-DMF, 140 °C, 63%) furnished the methyl ester of PBG lactam 14, a common intermediate in virtually all previous syntheses of 1. The present synthesis gives 14, which can be saponified to 1, in seven steps and 20% overall yield, representing a short and efficient route from commercially available reactants.

The direct alkylation of  $\beta$ -(hydroxymethyl)pyrroles is unprecedented, and several examples of the process (shown in the Table 1) illustrate its scope and generality.<sup>14</sup> Besides undergoing a two-carbon chain extension with dimethyl malonate, compound **12** reacts with NaCN to afford **15**, thus achieving a one-carbon homologation leading to nor-PBG **2**. Substituted malonates such as diethyl allylmalonate or diethyl benzamidomalonate also react with **12** to generate **16** and **17**, respectively, indicating the success of the method in forming quaternary carbon centers. Similar alkylations on the parent system, 3-(hydroxymethyl)pyrrole **18**, also give the expected products.

Pyrroles **16** and **17** represent clinically promising new directions for synthetic PBG analogs, both in PDT and other medical applications. Hydrolysis and decarboxylation of **16** afford allylated PBG **4**, which may find use in assembling haptens for the preparation of more avid anti-PBG antibodies. Such immunoreagents are currently employed to monitor serum PBG levels,<sup>15</sup> which are good indicators of lead poisoning in children.<sup>16</sup>

The synthesis of **17** illustrates an approach to new pyrrole-containing  $\alpha$ -amino acids like **5** of therapeutic interest in PDT. Since sophisticated active transport mechanisms for  $\alpha$ -amino acids have evolved in eucaryotic

 
 Table 1. Nucleophilic Alkylations of β-(Hydroxymethyl)pyrroles



<sup>*a*</sup> Representative conditions: 5 equiv of nucleophile, DMF, 130– 140 °C, 4 h. <sup>*b*</sup> 10 equiv used. <sup>*c*</sup> Combined yield for alkylation and deesterification–decarboxylation.

cells,<sup>17</sup> compounds like **17** may serve as precursors for highly tissue-selective PDT sensitizers, the attendant advantages of which include lower toxicity and reduced skin photosensitization.

**Acknowledgment.** We thank the National Institutes of Health for financial assistance in the form of a Program Project Grant, NIH GM 44874, in support of structure-based drug design. Support of the Cornell NMR Facility by the NSF (CHE 7904825; PGM 8018643) and NIH (RR02002) is gratefully acknowledged.

**Supporting Information Available:** Full experimental procedures as well as spectral and physical data are given for all new compounds reported (8 pages).

## JO961987J

<sup>(13)</sup> Related  $\alpha$ -alkylidenepyrrolenine cations have been invoked as intermediates in the formation of 2-alkylpyrroles from 2-(hydroxy-alkyl)pyrroles by reduction with lithium aluminum hydride or diborane: Biswas, K. M.; Jackson, A. H. *Tetrahedron* **1968**, *24*, 1145.

<sup>(14)</sup> One example of a related reaction, the alkylation of a  $\beta$ -[(*N*,*N*-dimethylamino)methyl]pyrrole with sodium dimethyl malonate, has been reported: see ref 7a.

<sup>(15)</sup> Gibbs, P. N. B.; Chaudhry, A. G.; Jordan, P. M. *Biochem. J.* **1985**, *230*, 25.

<sup>(16) (</sup>a) Rifai, N.; Cohen, G.; Wolf, M.; Cohen, L.; Faser, C.; Savory, J.; DePalma, L. *Ther. Drug Monit.* **1993**, *15*, 71. (b) Putnam, R. D. *Am. Ind. Hyg. Assoc. J.* **1986**, *47*, 700.

<sup>(17) (</sup>a) Meister, A.; Anderson, M. E. *Annu. Rev. Biochem.* **1983**, *52*, 711. (b) Christensen, H. N. *Biological Transport*, 2nd ed.; Benjamin: Reading, MA, 1975.